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Direct anti-metastatic efficacy by the DNA enzyme Dz13 and downregulated MMP-2, MMP-9 and MT1-MMP in tumours

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ABSTRACT

The DNA enzyme Dz13, targeted against the oncogene c-Jun, is capable of inhibiting various model tumours in mice albeit in ectopic models of neoplasia. In previous studies using orthotopic models of disease, the inhibitory effects of Dz13 on secondary growth was a direct result of growth inhibition at the primary lesion site. Thus, the direct and genuine effects on metastasis were not gauged. In this study, Dz13 was able to inhibit both locoregional and distal metastasis of tumour cells in mice, in studies where the primary tumours were unaffected due to the late and clinically-mimicking nature of treatment commencement. In addition, the effect of Dz13 against tumours has now been extended to encompass breast and prostate cancer. Dz13 upregulated the matrix metalloproteinase (MMP)-2 and MMP-9, and decreased expression of MT1-MMP (MMP-14) in cultured tumour cells. However, in sections of ectopic tumours treated with Dz13, both MMP-2 and MMP-9 were downregulated. Thus, not only is Dz13 able to inhibit tumour growth at the primary site, but also able to decrease the ability of neoplastic cells to metastasise. These findings further highlight the growing potential of Dz13 as an antineoplastic agent.

INTRODUCTION

Dz13 is a DNA enzyme designed originally to reduce intimal thickening in injured rat carotid arteries [1]. Since then, this particular ‘gene shear’ molecule has been shown to have potential therapeutic effects against a variety of disorders as mentioned below. DNAzymes are synthetic, single-stranded DNA-based catalysts engineered to bind to their complementary sequence in a target messenger RNA (mRNA) through Watson–

Crick rules for base-pairing and cleave the mRNA at predetermined phosphodiester linkages (reviewed in [2]). For example, Dz13 cleaves the target human c-Jun mRNA at position G¹³¹¹[1]. By way of a handful of critical studies, these biocatalytic molecules have emerged as a potential new class of nucleic acid-based drugs because of several beneficial attributes [2].

Dz13 has been shown in ectopic mouse tumour models to reduce the growth of melanoma indirectly via anti-angiogenesis [3], while exhibiting direct activity against squamous cell carcinoma [4], osteosarcoma, OS [5, 6] and liposarcoma [7]. In OS, this agent can be combined with a frontline drug such as doxorubicin for better efficacy [8], especially once it has been administered in a nanoencapsulated form [9]. While Dz13 has direct anti-tumour effects based on reduced cell growth and heightened cell death, the underlying mechanisms have not been elucidated.

The proteolytic breakdown of proteins of the extracellular matrix (ECM) has long been recognized as a hallmark of invading primary cancer lesions [10]. Several classes of proteases contribute to ECM breakdown and remodeling, most of which are upregulated in the course of metastatic cancer progression in different types of cancers [11]. Matrix metalloproteinases (MMPs) constitute a family of zinc-dependent endopeptidases that have been studied in the past few decades in the context of cancer, and the consensus view at present is that the main role of MMPs in angiogenesis, tumour growth and metastasis is degradation of ECM and release and/or activation of growth factors through such activity. Accordingly, inhibitors to MMPs have entered clinical testing, though the first clinical trials have led to disappointing results [12].

One of the critical proteases involved in cell migration is membrane-type 1 matrix metalloproteinase (MT1-MMP or MMP-14). MT1-MMP degrades extracellular matrix to furnish a path for cells to migrate, sheds cell surface molecules (that can serve as migratory signals), and activates extracellular signal-regulated protein kinase (ERK), thus enhancing cell migration [13]. Expression of MT1-MMP and activation of MMP-2 correlate with progression in human melanoma [14]. Relative risk of death in mesothelioma patients with low MT1-MMP expression is significantly lower than patients with high expression [15]. However, to date, the role of Dz13 against this MMP has not been evaluated.

The activation of the 72kDa type IV collagenase proMMP-2 (gelatinase A) correlates with increased occurrence of metastases, and leads to a conversion of the 72kDa pro-MMP species to a 63kDa zymogen, mediated by MT1-MMP activity at the cell surface [16]. In hepatic stellate cells, involved in liver healing, MT1-MMP activates MMP-13 which in turn activates MMP-9 [17]. In non-small-cell lung carcinoma, significant association with poor survival by both MMP-2 and MMP-9 has been reported [18]. Expression of MMP-2 and -9 is up-regulated in endometriomas and more pronounced in advanced stage disease [19]. In gastric cancer, expression of MMP-2 is strongly associated with tumour progression and lymph node metastasis [20].

Cleavage of the 92kDa type IV collagenase proMMP-9 (gelatinase B) results in its activated form, an 82kDa protein that has been reported to enhance the invasive phenotype of cultured MDA-MB231 cells due to increased capacity of degradation of ECM and transversing basement membrane following activation [21]. In epithelial ovarian cancer, overexpression of stromal MMP-9 and MT1-MMP is independently

associated with negative prognosis [22]. High-grade prostate tumours are more likely to express MMP-9 [23]. In uroepithelial carcinoma patients, increased pro-MMP-9 and active MMP-2 levels correlate with disease progression [24].

Thus, this study demonstrates that Dz13 can downregulate MMP-2, MMP-9 and MMP-14 levels in tumour cells, in addition to downregulating its target gene c-Jun, with no effect on the AP-1 transcription factor component (and associate of c-Jun), c-Fos. Moreover, this study seminally demonstrates the direct antimetastatic ability of Dz13 in models of tumour progression.

MATERIALS AND METHODS

Cells

The human prostate cancer PC3, breast cancer MDA-MB231, osteosarcoma SaOS-2, and osteosarcoma 143B cell lines were from the ATCC (Virginia, USA), while the osteosarcoma G292 cell line was from D. Thomas (Peter MacCallum Cancer Institute, Melbourne, Australia). Cells were propagated in α -MEM supplemented with 10% FBS and 1% antibiotic-antimycotic (complete medium). Cell lines were maintained up to 20 passages in a 37°C/5% CO₂ incubator, and were ensured to be >95% viable.

DNAzyme transfection

The 34mer, 10-23 class of deoxyribozymes, Dz13 and Scr (the scrambled sequence control for Dz13) oligonucleotides were synthesised and prepared according to established conditions [25]. DNAzyme (0.8 μ M) was transfected with Fugene-6 liposomes (Roche Diagnostics, Sydney, Australia) in complete medium.

Western blotting

Treated cell lysates were immunoblotted as per a published method [26]. Briefly, post-transfection, cells were gently lysed with RIPA (150mM NaCl, 50mM Tris, 1mM EDTA, 0.1% SDS, 1% Triton X-100 pH 7.4) buffer containing complete protease inhibitors (Roche Diagnostics). All primary antibodies were from Santa Cruz Biotechnology (Santa Cruz, CA, USA), and all horseradish peroxidase (HRP)-conjugated secondary antibodies were from Dako.

Immunohistochemistry (IHC)

IHC was performed as before [27] on paraformaldehyde-fixed paraffin-embedded tumour specimens. 5µm sections were deparaffinised in xylene, then rehydrated in a graded ethanol series. High pH (9.5) 10mM Tris/1mM EDTA buffer at 95°C was used for antigen retrieval. All primary antibodies were from Santa Cruz Biotechnology and were incubated overnight at 4°C. Secondary biotinylated antibodies (Dako, Sydney, Australia) were incubated with specimens at room temperature for 1h.

Establishment of metastasising 143B tumour model

Prior approval for all animal experimentation was obtained from the St. Vincent's Health Animal Ethics Committee. Female 5-week-old Balb/c nude mice ($n = 3$, sourced from the Animal Resources Centre, Perth, Australia) were anaesthetised with ketamine (100mg/kg) and xylazine (10mg/kg). 2.5×10^5 143B tumor cells in 50% Matrigel (BD Biosciences, Sydney, Australia) were injected into the inguinal fatpad of 5-week-old

female Balb/c nude mice. At the end of the study, the peritoneal cavity was checked for local metastases and the liver and lungs for distal metastases, photographs were taken, and tissues were processed histologically.

Proof of anti-metastatic activity of Dz13 against novel 143B tumour model

As above, 143B tumor cells in 50% Matrigel were injected into the inguinal fatpad of mice. Dz13 or Scr (250ng) in saline was administered intraperitoneally 6 weeks later when tumours became palpable. Four weeks later, primary tumours were weighed, and local and lung macrometastases were counted. The peritoneal cavity was checked for local metastases and the lungs for distal metastases, and tumours processed histologically.

Test of Dz13 against metastasis of cancer cells from an orthotopic tumour

Female 5-week-old Balb/c nude mice ($n = 4$ per group) were anaesthetised with ketamine (100mg/kg) and xylazine (10mg/kg). 2×10^4 SaOS-2 tumour cells in 50% Matrigel was injected intratibially in a volume of 20 μ L in the proximal tibia using a 26G needle and a gentle ‘screwing’ motion to prevent bone cortex rupture a before [28]. The general condition of all mice were observed before the tumours were measured apically (AP) and longitudinally (L) with digital callipers. Taking into consideration tumour bulge, AP was measured left-to-right across the knee-cap and L was an anterior-to-posterior measurement of the tibia (all measurements obtained where the tumour growth was maximal). Tibial tumour volume was calculated using the formula: $\frac{4}{3}\pi [0.25(AP+L)]^2$ [29]. A lung lobe of 3 representative mice from each treatment group was processed

histologically [30] and sectioned at 5µm at every 100µm interval. The number of micrometastases in each section was then counted for each cohort and summed.

Proof-of-principle Dz13 activity studies against ectopic tumors

Prior approval for use of mice was obtained from the St. Vincent's Health Animal Ethics Committee. Female 5-week-old Balb/c nude mice ($n = 5$ per group) were anaesthetised with ketamine (100mg/kg) and xylazine (10mg/kg). Dz13 was mixed with PC3, MDA-MB231 or SJS-1 tumour cells (1×10^6) at a concentration of 0.4µM in 50% Matrigel (BD Biosciences) prior to ectopic injection subcutaneously in the midback of 5 week-old Balb/C *nu/nu* mice. Backflow of the injectate was prevented by retracting the needle post-injection slowly from the injection site. Mice were fed and hydrated ad libitum and monitored twice weekly for tumour development, and then three times weekly when tumours were palpable. Tumour volumes were measured and tumours histologically processed as per published method [31, 32].

Statistical analysis

All data were analysed using the one-way student's *t*-test with unequal variances.

RESULTS

Locally and distally metastasising osteosarcoma model

In this study, when 143B cells were injected into the fatpad, palpable tumours arose at 3 weeks post-injection (**Fig. 1a**). At this early stage, tumours were almost white in colour

with a maximal dimension of 5mm across (**Fig. 1b**). Clearly discernible tumours appeared around 4 weeks (**Fig. 1c**). At the 6-week stage, tumours were approximately 1cm x 1cm in length and width dimensions, with a healthy pink hue and easily delineated vessels indicating aggressive tumour growth (**Fig. 1d**). At this point, several macrometastases were noted in the visceral walls surrounding peritoneal organs, including the intestines (**Fig. 1e**). Some of these locoregional growths were at least 1cm away from the parent tumour (**Fig. 1f**). Evidence of macrometastases was noted on the surface of lungs, though these were almost pin-point size. Confirmation with histology revealed that 143B cells did travel to and establish in the lungs from the peritoneum (**Fig. 1g**).

Dz13 directly inhibits local as well as distal tumour metastasis

The novel 143B model was then applied to directly test the effects of Dz13 against metastasis. Dz13 was administered 6 weeks after 143B cells were injected to ensure that primary tumours were unaffected (**Fig. 2a**) but an effect on metastasis could nevertheless be monitored. To this end, both local metastases in the peritoneal cavity and walls (**Fig. 2b**) and distal metastases to the lungs (**Fig. 2c**) were reduced. Thus, efficacy of Dz13 against metastasis was readily observed. The Scr control did not affect either primary tumour or metastasis. In addition, effects of Dz13 were tested on OS metastasis from the bone, this time with the established SaOS-2 tumour model [28]. Tumours were allowed to grow to around 1cm in both the AP and L dimensions, before Dz13 was administered intravenously in bolus dosages. Lung surface macrometastases were reduced in the Dz13 cohort of mice (**Fig. 3a**), and so were lung micrometastases (**Fig. 3b**). Histological

examination of lung lobe sections revealed that both size and number of micrometastases were reduced when animals were administered Dz13, but not Scr oligonucleotide (**Fig. 3c**). Tumour measurements at the tibiae of mice shows equal mean lesion volumes across the treatment groups (**Fig. 3d**).

Dz13 alters matrix metalloproteinase levels in tumour cells

Having demonstrated that Dz13 can directly inhibit tumour metastasis, the underlying molecular mechanisms were examined. Since MMP-2 and MMP-9 have been associated with c-Jun regulation [3, 4], these agents of ECM breakdown were explored. In addition, MT1-MMP (MMP-14) was also evaluated. Lysates for Dz13- and Scr-treated human osteosarcoma SJS-1 cells were immunoblotted. Levels of activated MMP-2 increased initially, but then decreased after 1h post-transfection with Dz13 (**Fig. 4**). Interestingly, MMP-9 was activated as time progressed post-transfection of cells with Dz13. However, for MT1-MMP, levels consistently decreased as the assay progressed.

Dz13 reduces ectopic osteosarcoma, prostate and breast tumour growths

Ectopic osteosarcoma (G292), prostate (PC3) and breast (MDA-MB231) tumour growths were reduced by Dz13 (**Fig. 5**). A statistical difference in tumour volume was noted at day 32 post-injection of cells, and at the end of the study (day 35), Dz13-treated tumour growths were clearly stunted. Less aggressive growth was noted with Dz13 treatment as manifested by sparse growth of tumour cells within the lesion (**Fig. 6**), which in itself was small and close to the underlying muscle. The Scr control tumours were the same volumes as the untreated tumours.

Dz13 downregulates target c-Jun and MMP levels in ectopic tumours

Ectopic osteosarcoma (G292), prostate (PC3) and breast (MDA-MB231) tumour sections were evaluated by immunohistochemistry for c-Jun and c-Fos (off-target control). c-Jun levels were decreased in all tumours, but c-Fos was not (**Fig. 7**). Likewise, MMP-2 levels were slightly lower in Dz13-treated tumours (**Fig. 8**). Immunohistochemistry for MMP-9 revealed a clear downregulation of this MMP as a result of Dz13 treatment of tumours (**Fig. 9**). Akin to results in vitro, levels of MT1-MMP were reduced in all tumours in vivo (**Fig. 10**).

DISCUSSION

In the future, Dz13 may be a feasible approach to tumour management. Metastasis is the event in tumorigenesis that signals advanced stage disease, and one that is unfortunately frequently not amenable to medical intervention. As such, agents capable of reducing the impact of metastasis are likely to significantly alter the current management strategies for cancers. This is surely the case for OS [33], where with good management strategies, the 10-year disease-free survival is about 60% in patients with localised disease and 30% in patients with metastatic disease at diagnosis.

c-Jun, in the form of the AP-I complex, regulates MMP-9 levels in mammalian cells [34]. This oncoprotein is also known to regulate expression of MMP-2 [35, 36]. In fact, in OS cells, c-Jun has been linked to both MMP-2 and MMP-9 activities and to cell ability to invade [37]. c-Jun also regulates MT1-MMP levels in mammalian cells [38]. As mentioned above, it has been previously demonstrated that Dz13, which

downregulates c-Jun, can in fact regulate MMP-2 expression and subsequent activity in endothelial cells [3] and MMP-2 and -9 in squamous cell carcinoma cells [4].

Thus, here we extend Dz13 action on MMPs, specifically showing that MT1-MMP is downregulated when OS cells are treated with Dz13, a novel finding since MT1-MMP was not evaluated in previous Dz13 studies [3, 4]. However, in discordance with previous findings, our studies show that Dz13 firstly upregulates MMP-2 in cultured OS cells, and levels decrease to baseline (undetectable) at later time-points. In addition, another variant from previous findings was the increasing MMP-9 activation in cells treated with Dz13 as time progressed. One explanation is that these findings could be due to the link that both MMP-2 and MMP-9 have with apoptosis [39], which is known to commonly occur with Dz13 treatment of tumour cells [40].

This study is also novel in that it seminally proved the direct effect of Dz13 on tumour metastasis. To test whether Dz13 can directly inhibit tumour metastasis, rather than as a result of primary tumour inhibition as demonstrated before [5-8], a locally metastasising model was established. 143B cells were injected into the inguinal fatpad of mice, at a region which facilitates easy manipulation of the mice (drug administration and hence testing without anaesthesia of animals, site at which animal cannot physically affect tumour growth with limbs or snout, tumour measurements can be obtained without need for animal immobilisation, and cells are secured in a specified spot within fatpad and not floating around in peritoneum). OS does in fact metastasise to the peritoneum in patients, albeit from organs other than bone [41, 42]. One advantage of this model over other OS models using human cells [28, 43] is the presence of both regional and distal metastases in the peritoneum and surrounding walls and the lungs respectively. This model can be

used for testing of the antimetastatic properties of candidate therapeutic agents against OS due to the aforementioned advantages.

In both the 143B fatpad model and in an orthotopic model for OS, where cells are injected directly into bone [28], primary tumours were left to grow until a stage where treatment would have no effect on the primary lesion. When Dz13 was then used, the development of metastases was significantly inhibited in both models. The Scr control oligonucleotide had no effect on tumours.

While MMPs levels were perturbed in cultured cells, the effects of Dz13 *in vivo* needed to be tested in a panel of tumour types. The tumours evaluated consisted of prostate, breast and osteosarcoma, though ectopic models were used to examine protein levels via tumour section immunohistochemistry (IHC). When IHC was performed, a consistent decrease in MMP-2 and MMP-9 levels was noted in the Dz13 cohort of animals compared to both the saline-treated and Scr-treated tumours. Thus, while *in vitro* results suggest that MMP levels may fluctuate and even increase via activation, *in vivo* results showed a consistent decrease in MMPs by Dz13. The discrepancy could be due to the fact that in culture, cells were exposed to a very potent Dz13 onslaught in a 2-D configuration, while *in vivo*, cells were protected by the Matrigel and the 3-D nature of the injected 'colony' of cells, and treatment was protracted over 5 weeks. However, what was consistent was the finding that both *in vitro* and *in vivo*, levels of MT1-MMP were reduced in tumour cells treated with Dz13 but not the scrambled control.

In addition, *in vivo*, significant decrease in c-Jun levels were noted for all ectopic tumours treated with Dz13 but not when cells were treated with the Scr control oligonucleotide. *In vitro*, cells have to be serum-starved prior to growth induction (and c-

Jun elevation) before Dz13 shows its effects [1, 3, 4], but in vivo, where such a protocol is irrelevant, except in areas of the tumour where new blood supply has been established after a certain degree of ischaemia, Dz13 is active against its target mRNA, the c-Jun oncogene. No changes in c-Fos levels were noted. Thus, for DNAzyme technology in general, the need for serum-starvation in culture [31, 32], while providing critical proof that the catalytic nucleic acid is active against its target, may fail to be indicative of what occurs in vivo, and in fact is an underestimation of the potent action of Dz13 against its target.

For Dz13, the present set of results further highlight the inherent potential of this molecule. Not only is it able to control tumour at the primary site, but also at the secondary site as well. As for other tumours, and particularly in OS, while the primary tumour may be removed in a good number of cases, it is the metastases which become life-threatening [44]. Further studies with other metastasising tumours may prove the widespread beneficial effects of Dz13.

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AUTHOR CONTRIBUTIONS

CRD conceived the project and designed the experiments. CRD and MLT performed the studies. PFMC provided clinical foresight to the studies. CRD prepared, and MLT and PFMC edited the manuscript. CRD gave approval for the final version to be submitted.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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CAPTIONS TO FIGURES

Figure 1. Characterisation of the 143B locally metastasising model. This model can be used for testing of the antimetastatic properties of candidate therapeutic agents against osteosarcoma. 143B cells were injected into the inguinal fatpad of mice. A, palpable tumour at 2 weeks post-injection, B, exposed tumour at 2 weeks post-injection, C, clearly discernible tumour at 4 weeks, D, exposed tumour at 4 weeks, E, *white arrowhead* indicates site of primary tumour, *black arrowhead* indicates site of secondary locoregional tumour, F, *arrowhead*, site of locoregional tumour metastasis, G, *arrowhead*, pulmonary metastasis within lung parenchyma. *Scale bar = 25µm*.

Figure 2. Dz13 inhibits 143B tumour metastasis. Dz13 was tested in the novel intraperitoneal 143B model. (A) Primary tumours were unaffected. (B) Dz13 inhibits

tumour metastasis locally in the peritoneum. (C) Dz13 inhibits tumour metastases to a distal site (lung). $n = 5$, error bars represent standard deviations.

Figure 3. Dz13 inhibits OS metastasis from the bone efficaciously. Dz13 efficacy was next tested in the SaOS-2 orthotopic model. Dz13 was able to reduce OS metastasis to the lung after treatment was administered when tumours were palpable. Both macrometastasis (A) and micrometastasis (B) counts were reduced in the Dz13 cohort. (C) Histology confirmed that there were less micrometastases in lungs of mice in the Dz13 cohort (middle) compared to untreated (top) and Scr-treated (bottom). (D) Primary tumour was not affected by the treatments administered.

Figure 4. Dz13 downregulated matrix metalloproteinases in tumour cells. Two osteosarcoma cell lines were transfected with Dz13 and cell lysates harvested at the indicated time-points. Lysates were immunoblotted for the MMP-2, -9 and -14 (MT1-MMP). GAPDH indicates equivalent loading between samples.

Figure 5. Dz13 reduces volume of ectopic (subcutaneous) osteosarcoma, prostate and breast tumour growths. Ectopic osteosarcoma (G292), prostate (PC3) and breast (MDA-MB231) tumour growths were reduced by Dz13. Real-time tumour growth graphs are presented. Inserts show photographs of harvested representative tumours from the cohorts for each cell line. $n = 5$, error bars represent standard deviations.

Figure 6. Dz13 reduces aggressiveness of ectopic osteosarcoma, prostate and breast tumour growths. Haematoxylin- and eosin-stained osteosarcoma (G292), prostate (PC3) and breast (MDA-MB231) tumour sections are shown. In the Dz13 cohorts, less aggressive tumour growth is noted as sparse cell density and smaller area of occupation. *Arrowheads: black*, areas of dense cell growth, *white*, areas of sparse cell growth. *Scr*, scrambled sequence oligonucleotide of Dz13. *Scale bar = 25µm*. Representative images are shown. *Scale bar = 25µm, n = 5*.

Figure 7. Dz13 downregulates target c-Jun levels in ectopic osteosarcoma, prostate and breast tumours. Ectopic osteosarcoma (G292), prostate (PC3) and breast (MDA-MB231) tumour sections were evaluated by immunohistochemistry for c-Jun and c-Fos (off-target control). c-Jun and c-Fos (inserts)-immunostained tumour sections are shown. Representative images are shown. Negative control sections (no primary antibody) ruled out non-specific staining. *Scale bar = 25µm, n = 5*.

Figure 8. Dz13 downregulates MMP-2 in ectopic osteosarcoma, prostate and breast tumours. Ectopic osteosarcoma (G292), prostate (PC3) and breast (MDA-MB231) tumour sections were evaluated by immunohistochemistry for MMP-2. Representative images are shown. Negative control sections (no primary antibody) ruled out non-specific staining. *Scale bar = 25µm, n = 5*.

Figure 9. Dz13 downregulates MMP-9 in ectopic osteosarcoma, prostate and breast tumours. Ectopic osteosarcoma (G292), prostate (PC3) and breast (MDA-MB231)

tumour sections were evaluated by immunohistochemistry for MMP-9. Representative images are shown. Negative control sections (no primary antibody) ruled out non-specific staining. *Scale bar = 25µm, n = 5.*

Figure 10. Dz13 downregulates MT1-MMP in ectopic osteosarcoma, prostate and breast tumours. Ectopic osteosarcoma (G292), prostate (PC3) and breast (MDA-MB231) tumour sections were evaluated by immunohistochemistry for MT1-MMP. Representative images are shown. Negative control sections (no primary antibody) ruled out non-specific staining. *Scale bar = 12.5µm, n = 5.*

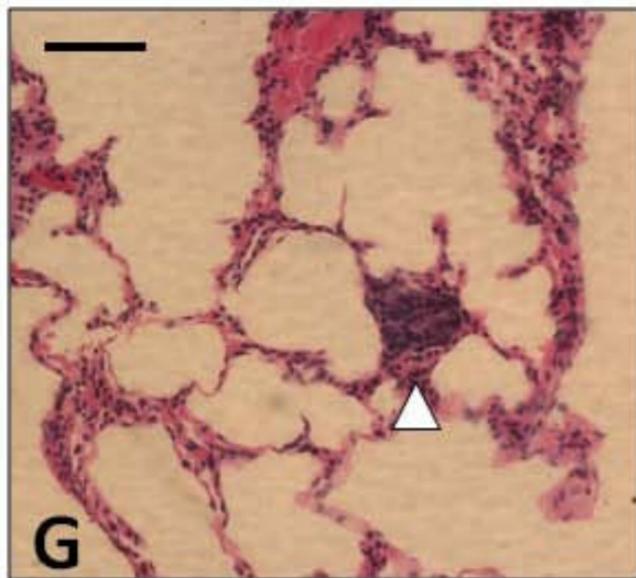
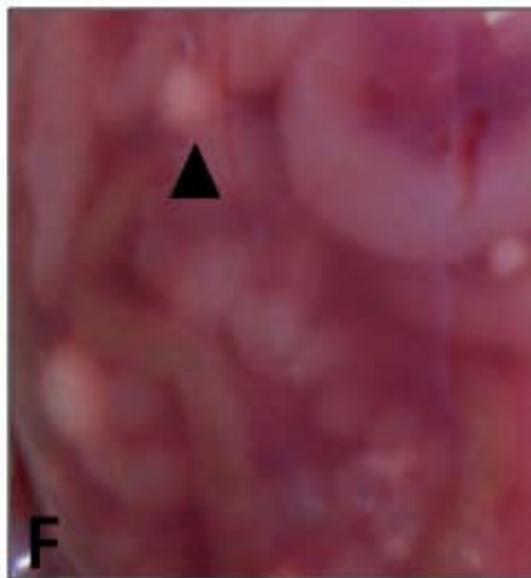
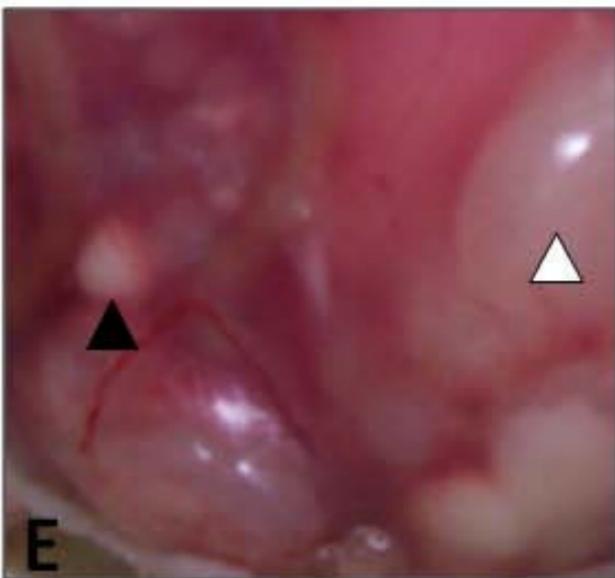
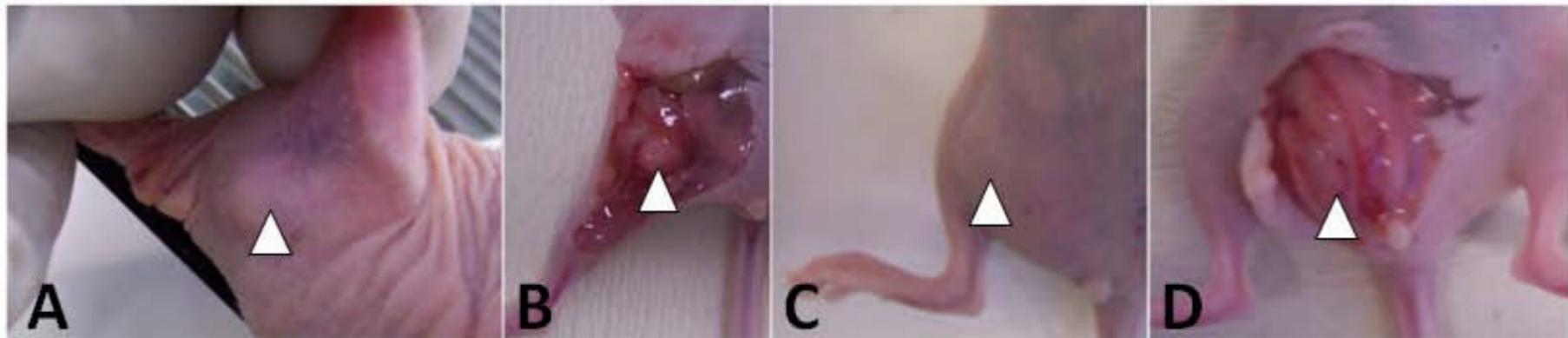


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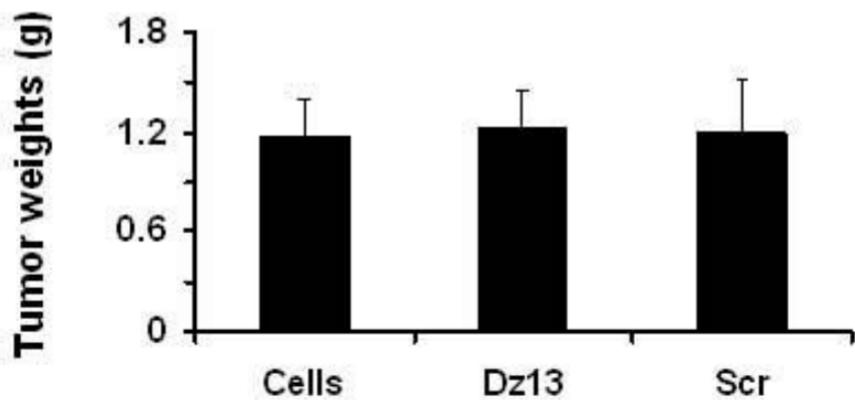
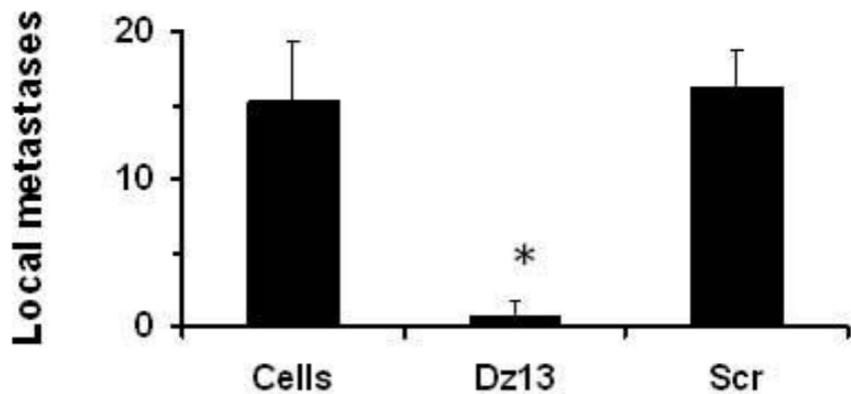
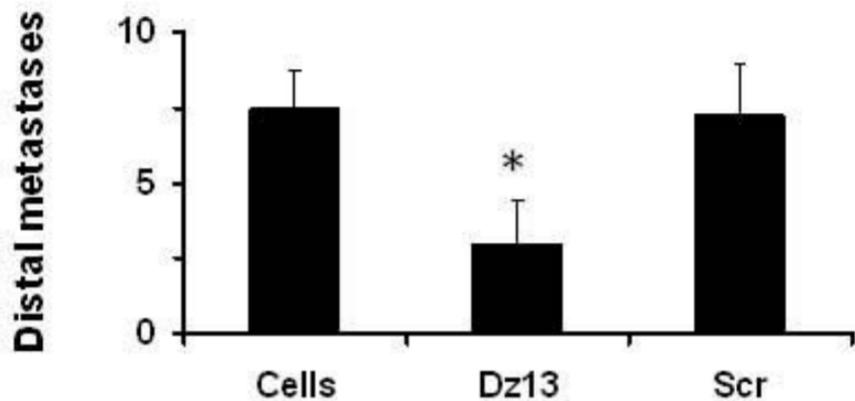
A**B****C**

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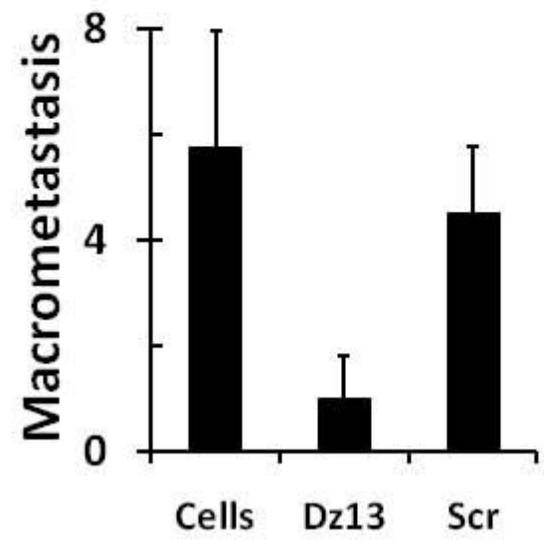
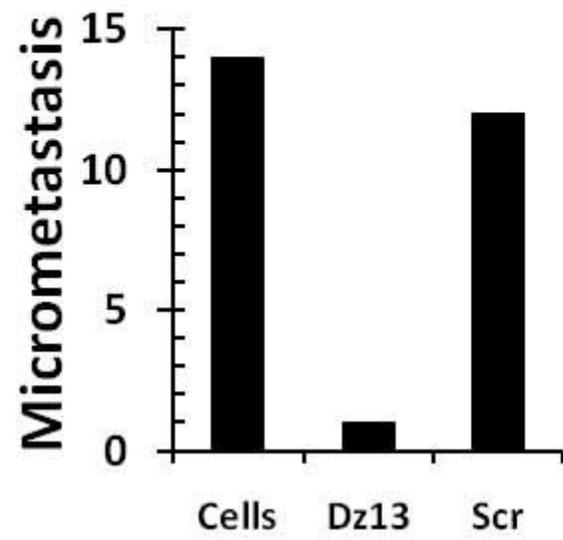
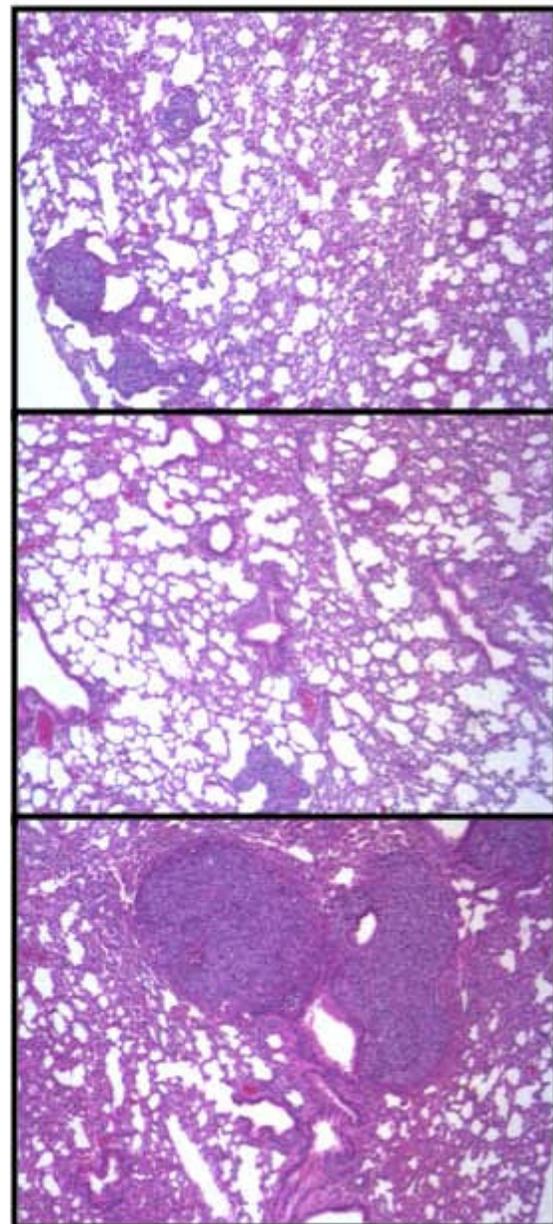
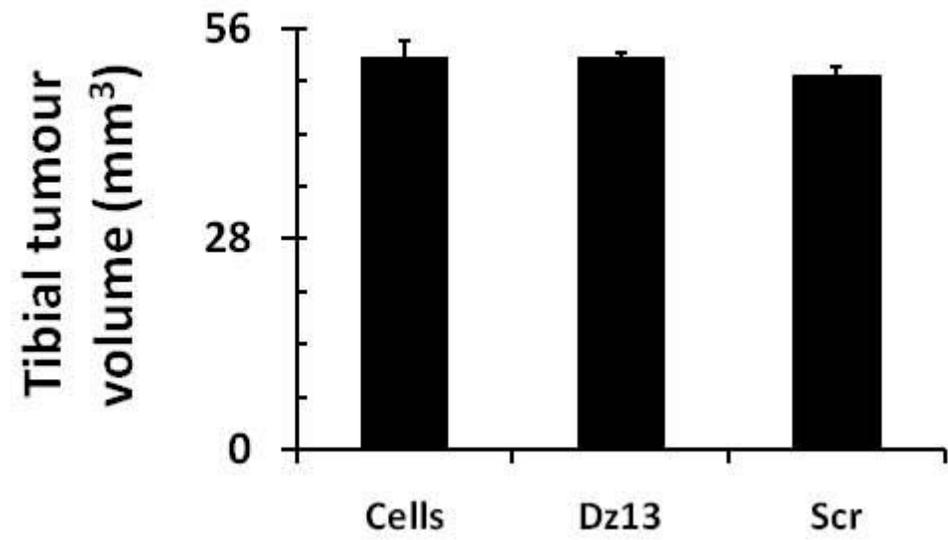
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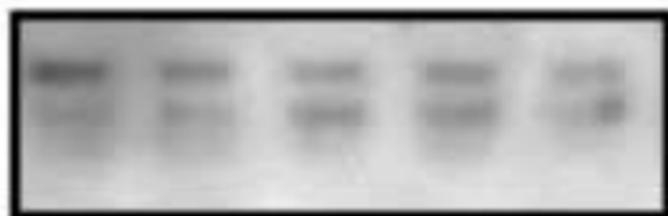
Time (h) 0 0.5 1 2 7

MMP-2



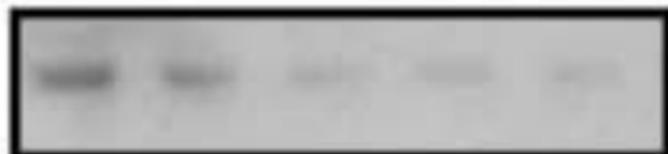
72kDa
63kDa

MMP-9



92kDa
82kDa

MT1-MMP



63kDa

GAPDH



37kDa

Figure 4

Tumour volume (mm³)

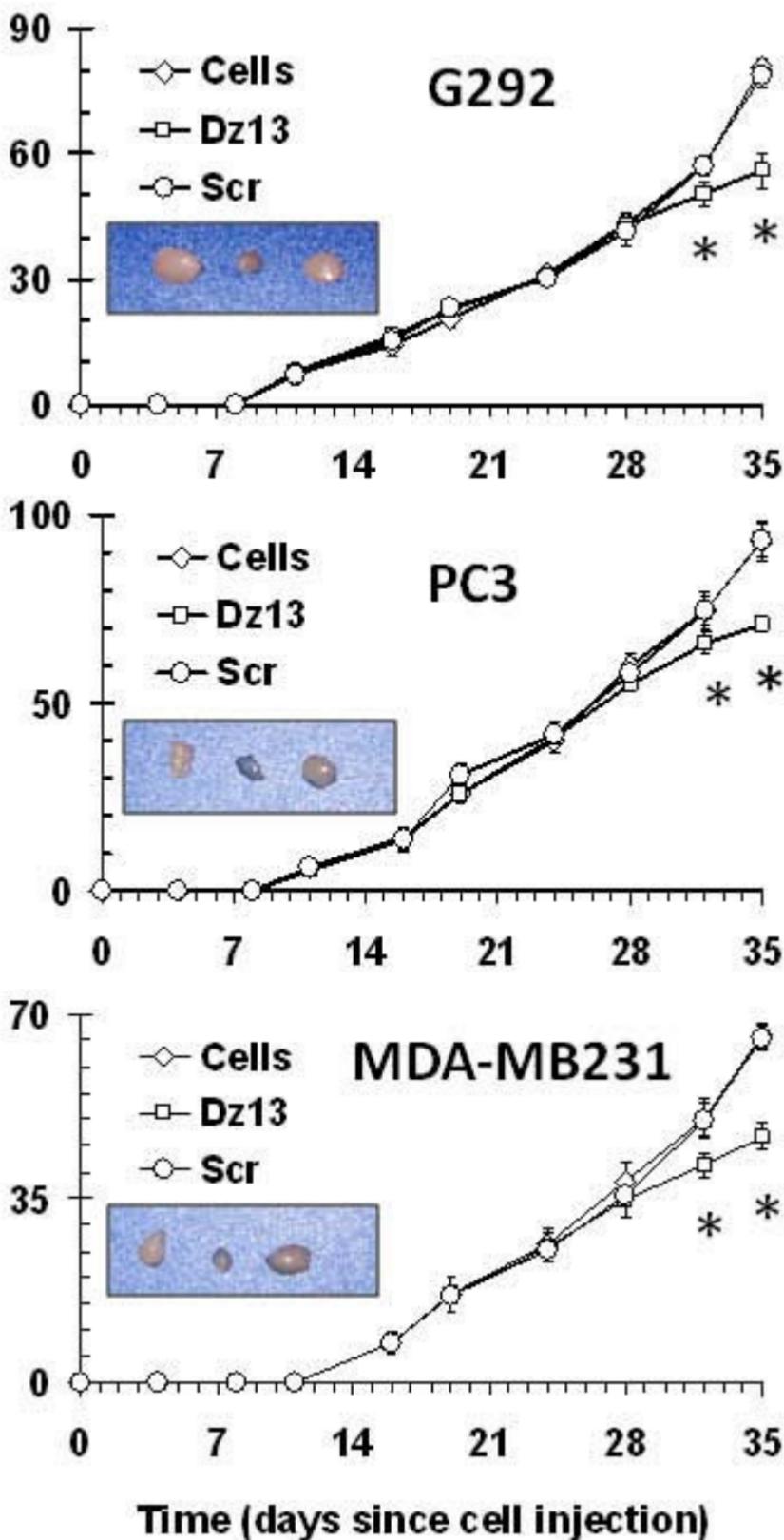


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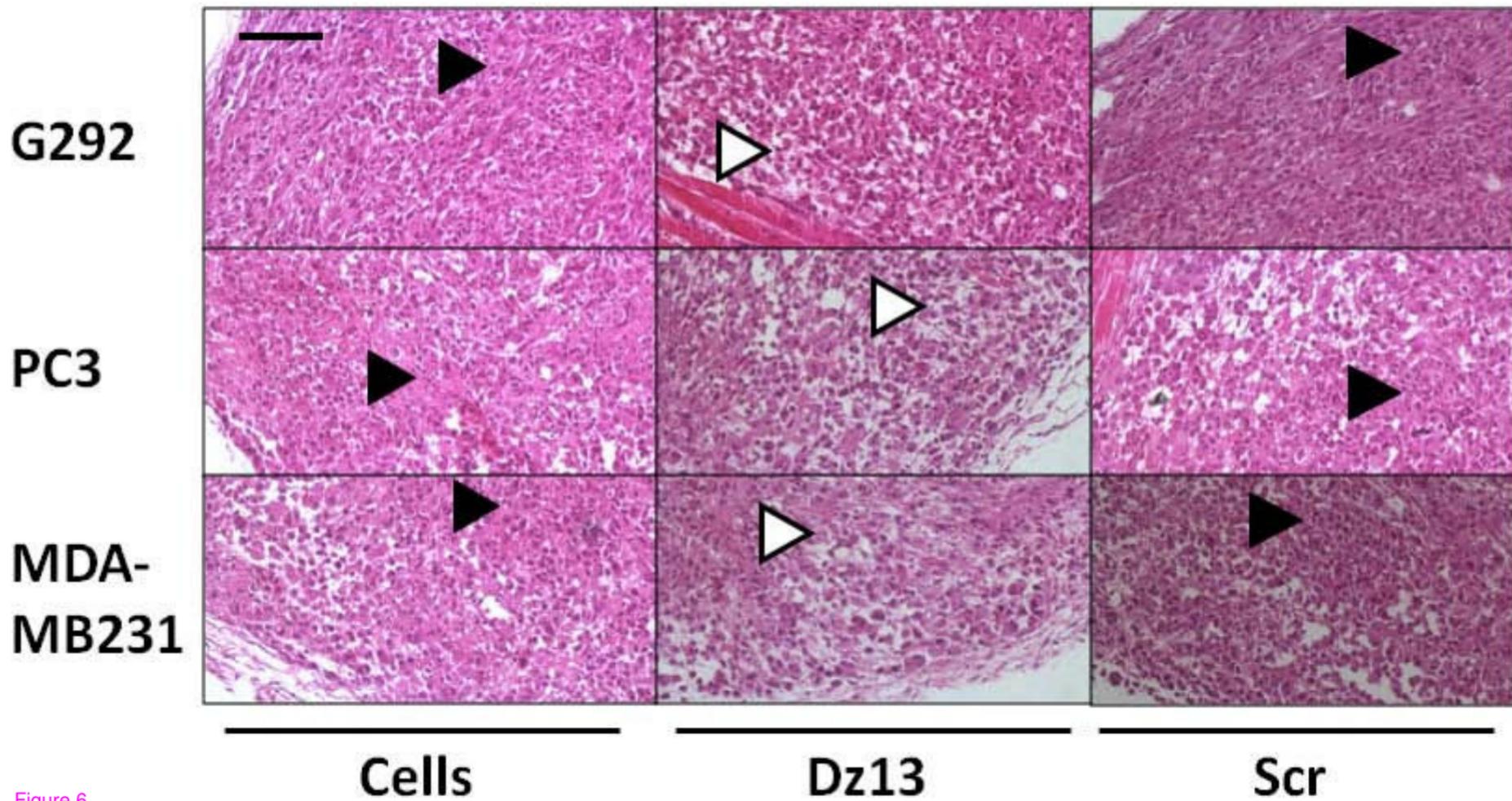
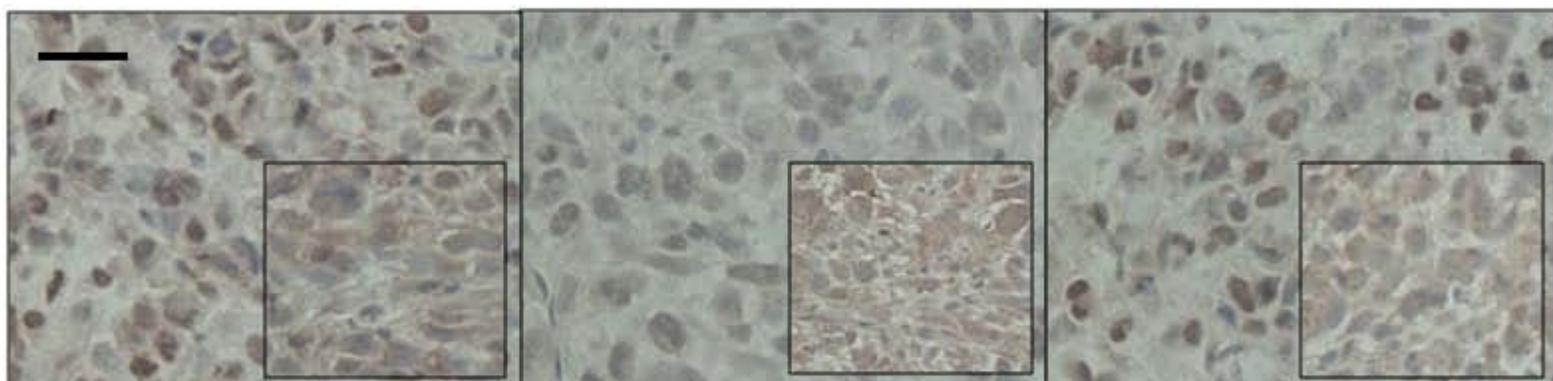
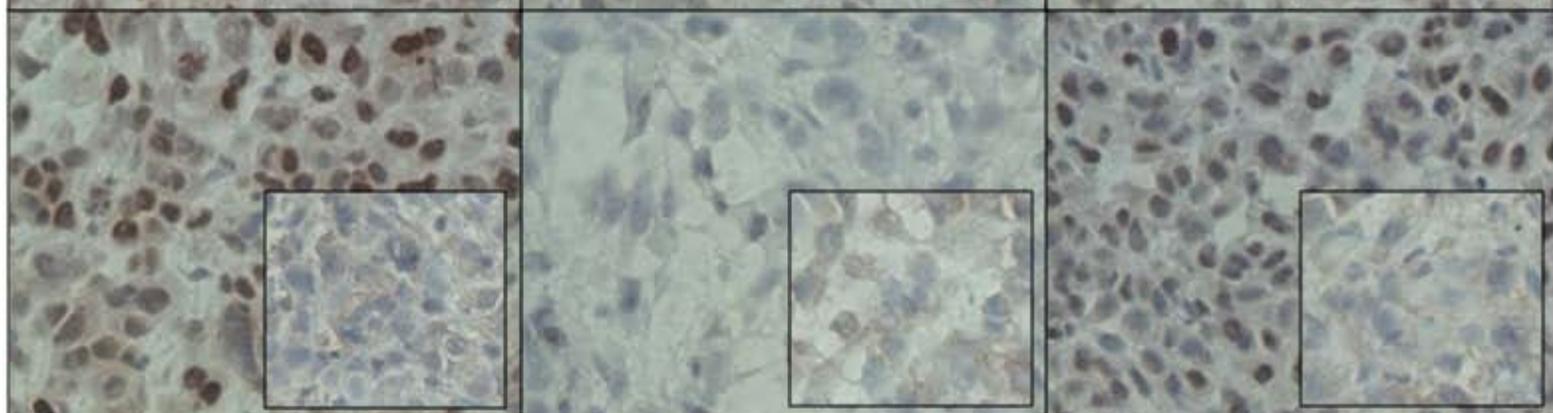


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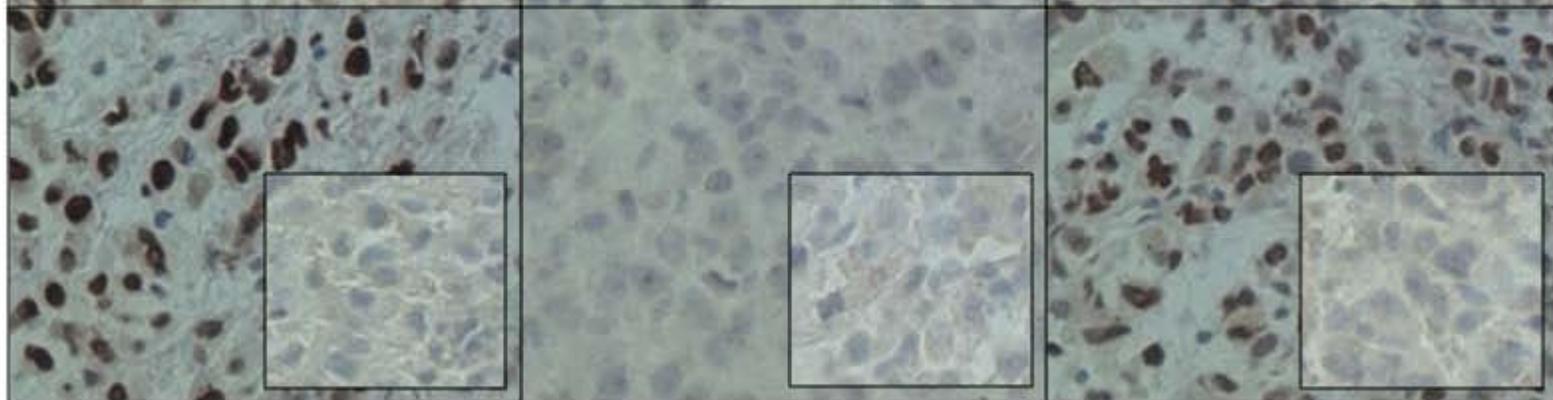
G292



PC3



**MDA-
MB231**



Cells

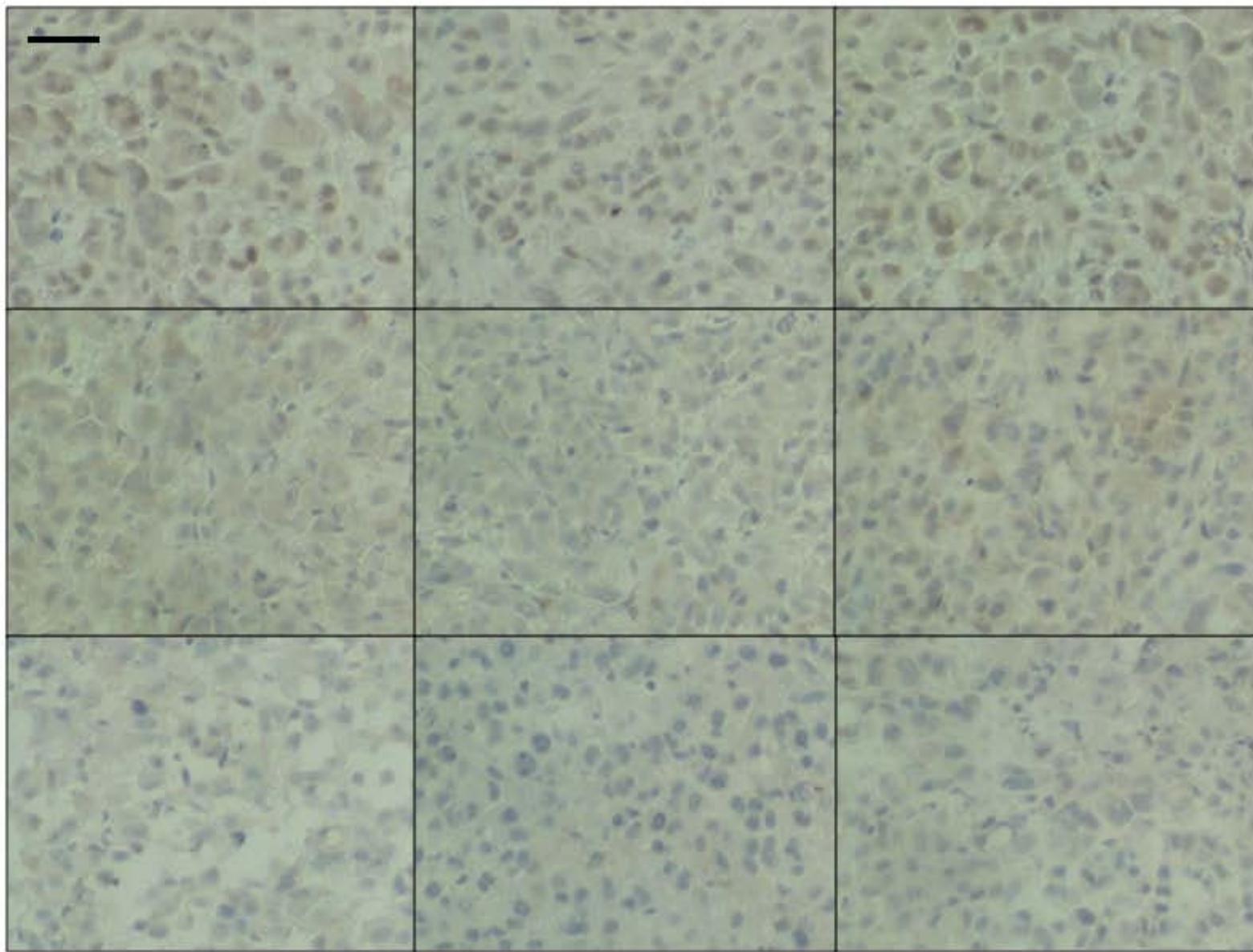
Dz13

Scr

G292

PC3

**MDA-
MB231**



Cells

Dz13

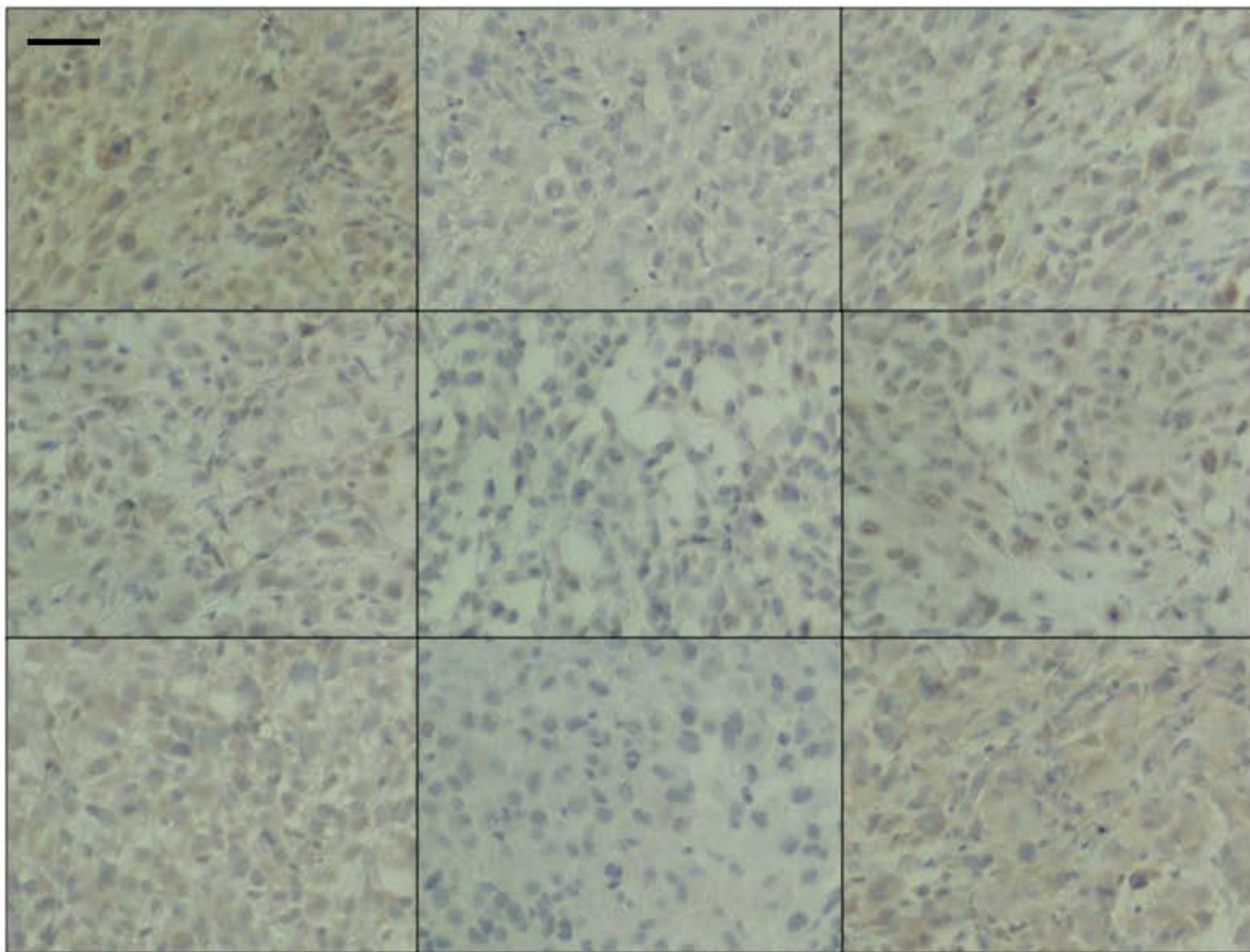
Scr

Figure 8

G292

PC3

**MDA-
MB231**

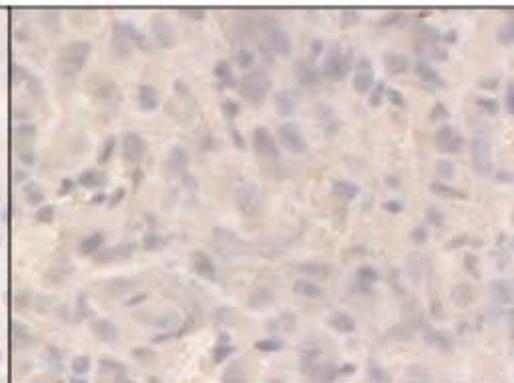
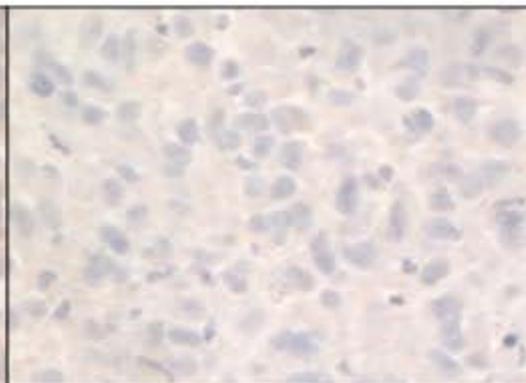
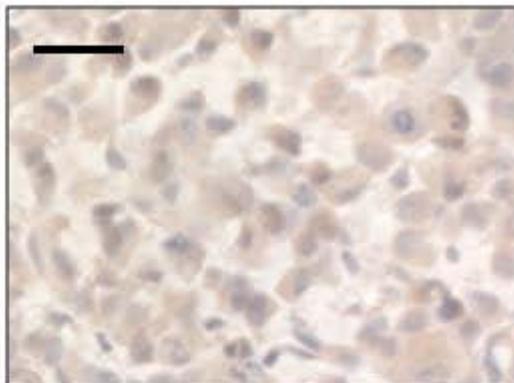


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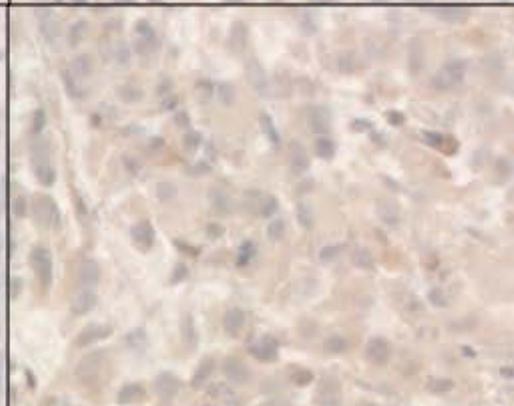
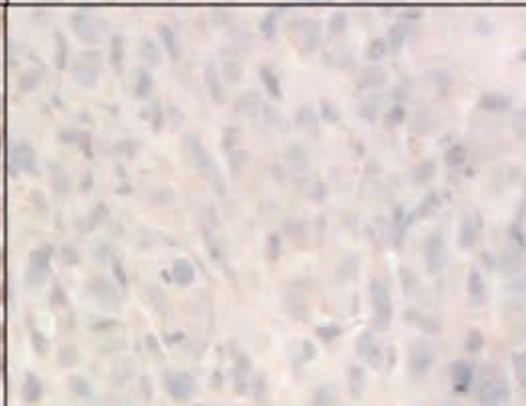
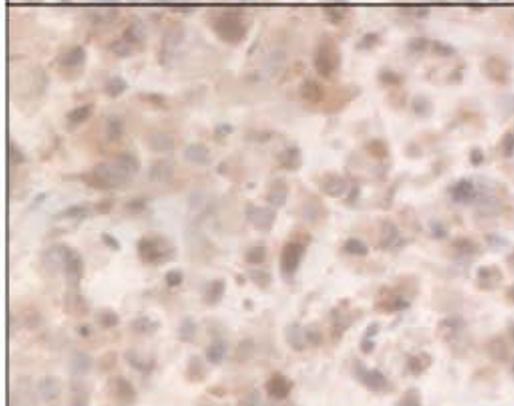
Dz13

Scr

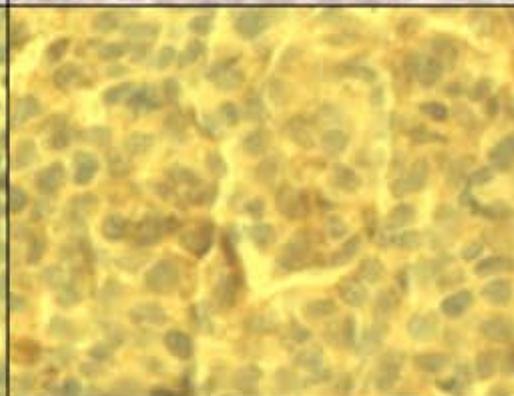
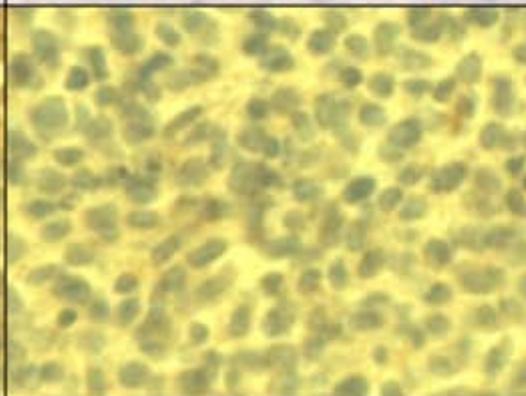
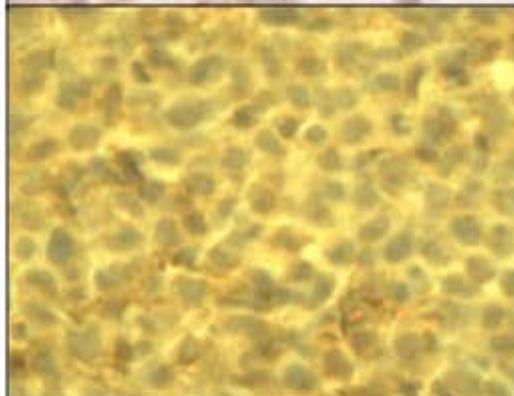
G292



PC3



**MDA-
MB231**



Cells

Dz13

Scr